

Seasonality, parity rates and transmission indices of *Mansonella ozzardi* (Manson) (Nematoda: Onchocercidae) by *Cerqueirellum argentiscutum* (Shelley & Luna Dias) (Diptera: Simuliidae) in a lower Solimões River community, Amazonas, Brazil¹.

Jansen Fernandes de MEDEIROS², Victor PY-DANIEL²

ABSTRACT

Mansonella ozzardi is transmitted by two dipterian families, Ceratopogonidae (midges) and Simuliidae (black flies). In Brazil, black flies are vectors for this filariasis until now. In this paper, we determined the seasonality, parity capacity and parasitic infection rate of *Cerqueirellum argentiscutum*. The work was carried out in the Porto Japão community, Lower Solimões River, Amazonas, Brazil. Results show that the black flies were more abundant during the rainy season (from December to May). The number of parous flies was higher in every sampling during the course of year. Monthly Biting Rate (MBR₁ 123742.00, MBR₂ 86701.50) was high, although Parasitic Infection Rate (PIR₁ 0.06, PIR₂ 0.08) and Annual Transmission Potential (ATP 7.25) were low in numbers.

KEY WORDS

Simuliidae, *Mansonella ozzardi*, Amazônia, Brazil.

Sazonalidade, taxa de paridade e índices de transmissão de Mansonella ozzardi (Manson) (Nematoda: Onchocercidae) por Cerqueirellum argentiscutum (Shelley & Luna Dias) (Diptera: Simuliidae) em uma comunidade do baixo rio Solimões, Amazonas, Brasil.

RESUMO

Mansonella ozzardi é transmitida por dois grupos distintos de insetos, Ceratopogonidae e Simuliidae. No Brasil, os simulídeos são os vetores dessa filariose. Neste trabalho, o objetivo foi determinar a sazonalidade, a paridade e a taxa de infecção parasitária de *Cerqueirellum argentiscutum*. O experimento foi realizado na comunidade Porto do Japão, Baixo rio Solimões, Amazonas, Brasil. Os simulídeos foram mais abundantes no período de chuva (dezembro a maio). Em todos os meses de coletas o número de paríparas foi maior. A taxa mensal de picada (TMP₁ 123742.00, TMP₂ 86701.50) foi elevada, já a taxa de infecção parasitária (TIP₁ 0.06, TIP₂ 0.08) e o potencial de transmissão anual (PTA 7.25) foram considerados baixos.

PALAVRAS-CHAVE

Simuliidae, *Mansonella ozzardi*, Amazônia, Brasil.

¹This work is part of the first author's PhD Thesis

² Instituto Nacional de Pesquisas da Amazônia, Coordenação de Pesquisas em Ciências da Saúde, Laboratório de Filarioses e Vetores, C.P. 478, 69011-970, Manaus, AM, Brasil, E-mail: jmedeiro@inpa.gov.br, pydaniel@inpa.gov.br

INTRODUCTION

Mansonella ozzardi is the filarial agent of mansonelliasis. The first report of this disease was made by Manson in 1897, in Guiana. According to Batista *et al.* (1960) and Oliveira (1961), people infected with mansonelliasis and high microfilaremia present symptoms of moderate fever, articular pain, adenite (followed by dizziness) and headaches. A new symptomatology attributed to this filariasis is the occurrence of visual lesions, which in turn can lead to blindness (Branco *et al.*, 1998, Garrido & Campos, 2000).

This filariasis is found from Mexico to Argentina, Guatemala and Panama in Central America, and also some islands of the Antillean archipelago. Except for Chile, Uruguay and Paraguay, every country in South America has already reported the presence of this parasite (Tavares & Fraiha Neto, 1997).

M. ozzardi is transmitted by insects of the families Ceratopogonidae and Simuliidae, Diptera. Initial work accomplished by Buckley (1934) indicated *Culicoides furens* (Poey) (Ceratopogonidae) as the vector of *M. ozzardi* in St Vincent Island, Caribbean. *C. furens* was later revealed as a vector in Mexico and Trinidad (Biagi *et al.*, 1958), whereas *C. phlebotomus* (Williston) was the vector in Haiti (Nathan, 1978, Lowrie & Raccurt, 1981).

In Colombia, *Simulium sanguineum* [= *Cerqueirellum sanguineum* (Knab)], *S. amazonicum* [= *C. amazonicum* (Goeldi)], *S. argentiscutum* [= *C. argentiscutum* (Shelley & Luna Dias)] and *Culicoides insinuiatus* Ortiz & Leon have been reported as likely vectors of *M. ozzardi* (Tidwell *et al.*, 1980, Tidwell & Tidwell, 1982). *S. sanguineum* [= *C. sanguineum* (Knab)], *S. minusculum* [= *Psaroniocompsa incrustata* (Lutz)] and *S. sanchezi* [= *C. oyapockense* (Floch & Abonnenc)] were indicated as vectors of *M. ozzardi* in south Panama, Guyana and Venezuela, respectively (Nathan *et al.*, 1982, Peterson *et al.*, 1984, Yarzabal *et al.*, 1985). Shelley & Coscarón (2001) observed in *S. exiguum* [= *Notolepria exigua* (Roubaud)] and *Culicoides labillei* (Iches) the microfilarial development of *M. ozzardi* to the infective stage (L₃) in (northern) Argentina.

Early assessment by Cerqueira (1959) indicated *S. amazonicum* [= *C. amazonicum*] as a vector of *M. ozzardi* in Brazil. However, Moraes *et al.* (1985) suggested that the species identified in that work was probably *S. argentiscutum* [= *C. argentiscutum*]. Shelley & Shelley (1976) also reported *S. amazonicum* [= *C. amazonicum*] as a vector of *M. ozzardi*, while Shelley *et al.* (1980) indicated not only *S. amazonicum* [= *C. amazonicum*], but also *S. argentiscutum* [= *C. argentiscutum*] involvement in the transmission of mansonelliasis. Moraes *et al.* (1985) indicated *S. oyapockense* [= *C. oyapockense*] as a vector in the State of Roraima, Brazil.

In Brazil, the first report of *M. ozzardi* was made by Deane (1949). Lacerda & Rachou (1956) provided evidence that this filaria is found in the State of Amazonas, in the communities bordering the Solimões River and its tributaries.

The goal of this work is to assess the seasonality, the parity and the transmission indices of *C. argentiscutum* one of the vectors of *M. ozzardi* in Brazil. This is the first work considering the range of a 1-year sampling of *C. argentiscutum*. This work follows the nomenclature according to Py-Daniel & Moreira Sampaio (1994), where the subgenus is elevated to genus level in Simuliidae.

MATERIALS AND METHODS

Study area

This work was conducted from September 1999 to August 2000, in the Porto Japão community (3°34" S / 61°09" W), lower Solimões River, Amazonas, Brazil. The community has approximately 100 people, subsisting on fishing, hunting and farming.

Procedure of Capture

The black flies were collected monthly during four consecutive days starting at 6:00 and ending at 18:15. The sampling intervals were divided into 15-minute periods, each followed by 15-minute intervals. They were captured by a laboratory technician, using manual suction collectors, according to the methodology of Medeiros & Py-Daniel (1999) and Medeiros & Py-Daniel (2003).

Analysis of the Ovaries

Between samplings, we analysed the ovaries to verify whether the females were nulliparous or parous, according to the methodology of Ramirez-Perez (1977). The flies were then placed in tubes containing 70% ethanol, for later examination at the Laboratório de Filarioses e Vetores of the Instituto Nacional de Pesquisas da Amazônia, Manaus, Brazil.

Dissection of the black flies

The flies were immersed in acid hematoxylin and stained for a period of 48 hours, prior to dissection under a stereomicroscope. They were placed on a slide, divided in three parts (head, thorax and abdomen), placed in a drop of a glycerin and covered by a coverslip. The slides were then observed by light microscopy, and the flies were verified for mansonelliasis infections. When the parasites were found, the developmental stages (mf, L1, L2, L3) were identified according to the method of Yarzabal *et al.* (1985).

Parasitic Infection Rate (PIR)

Two methods were used to obtain this index: PIR₁, considered the number of females infected by any filarial stage (mf, L1, L2 and L3) of *M. ozzardi* divided by the number of parous females

and dissected nulliparous X 100; PIR_2 considered the number of females infected with any filarial stages, except microfilaria (L1, L2 and L3) of *M. ozzardi* divided by the number of parous females dissected X 100 (Py-Daniel *et al.*, 2000).

Monthly Biting Rate (MBR)

This index estimates the number of black flies that bite a person exposed in the same place for a month, subjected to bites all day long (Davies & Crosskey, 1991). Two ways of calculating MBR were taken into account: one considers the total of captured females (nulliparous + parous) while the other considers only the parous females (Py-Daniel *et al.*, 2000).

$$MBR_1 = \frac{\text{Total number of females caught} \times N^0 \text{ of days in a month}}{\text{Number of catching days}}$$

$$MBR_2 = \frac{\text{Total number of parous females caught} \times N^0 \text{ of days in a month}}{\text{Number of catching days}}$$

Monthly Transmission Potential (MTP)

This index estimates the number of L3 stage larvae that can be transmitted to a person exposed to the vector during a one month period (Duke, 1968, Davies & Crosskey, 1991).

$$MTP = \frac{MBR_1 \times \text{Number of L3 stage larva by black flies}}{\text{Total number of black flies dissected}}$$

Annual Transmission Potential (ATP)

represents the number of L3 stage larvae that can be transmitted to a person exposed to the vector during a one year period (Davies & Crosskey, 1991). It is calculated by the sum of all MTPs (Monthly Potential Transmission) (Py-Daniel *et al.*, 2000).

Prevalence

Approximately 80% of the habitants in Porto Japão were examined. A lancet was used to puncture the fourth finger of individuals and a drop of blood was used to make a thick film on a microscope slide. The film was de-haemoglobinized, fixed in 80% methanol, and later stained with Giemsa and examined for microfilariae.

Additional Information

Ribeiro & Adis (1984) consider two seasons for the Central Amazon, a dry season between June and November and a rainy season from December to May.

The statistical analyses were performed as non-parametric tests, using the Mann-Whitney (U-test).

RESULTS

Seasonality

16320 *C. argentiscutum* females were collected throughout this work, with an average of 1360 individuals per month and 340 individuals per day. The highest number of simuliid flies was obtained during the rainy season (from December to May), with 13488 (82.65% of the total collected flies) (2248 ± 1373.9 flies/month / 562 ± 366.3 flies/day), showing a statistically significant difference ($U = 325$ $P < 0.001$) in relation to the number of flies collected in the dry season (from June to November) with a total of 2832 individuals (472 ± 305.4 flies/month / 118 ± 86.3 flies/day), approximately 17.35% of the total (Table 1).

During the rainy season, the highest incidence of simuliid flies was reported from December 1999 to February 2000, resulting in a total of 9753 specimens (59.76% of the total/year), (3251 ± 1290.2 flies/month / 813 ± 363 flies/day). In the dry season, the highest number of black flies was obtained in September/1999, November 1999 and June 2000, resulting in a total of 1994 individuals (8.53% of the total/year) (664.6 ± 306 flies/month / 166.1 ± 94 flies/day).

The highest number of individuals was collected in January (1021.5 ± 239.3 flies/day), corresponding to approximately 25% of the total. The lowest number of individuals was sampled in July (23 ± 18.3 flies/day), corresponding to approximately 0.56% of the total flies collected throughout this study.

Ovarian physiological stage

A total of 11441 parous females (70.1%) and 4879 nulliparous females (29.9%) were collected. The abundance of parous flies over the nulliparous flies was verified in every month of the year (Figure 1).

Parasitic Infection Rate - PIR

Only nine out of the 16.320 collected flies were infected by *M. ozzardi*, representing a Parasitic Infection Rate (PIR_1) of 0.06%, and a PIR_2 of 0.08%. The highest number of infected simuliid flies was found in January and February. The highest PIRs was reported in July (Table 2).

A total of 34 *M. ozzardi* larval stage was found in the black flies collected, with a higher number of L2 stage (17), followed by L1 (16) and L3 (1). All the larval stages of *M. ozzardi* were found in the thorax of the flies.

Monthly Biting Rates – MBR, Monthly Transmission Potential - MTP and Annual Transmission Potential - ATP

The estimated Monthly Biting Rate (BMR_1) was 124.521,25 bites/person-month and BMR_2 was 87.348,75. Both BMR_1 and BMR_2 were higher in January (31.674,25) and February (30.232,75) while lower BMRs were reported in July (690,0) and August (2.549,75) (Table 2).

The MTP was only calculated for February, when a L3 stage was found in an infected black fly. Therefore, the calculated MTP has the same value as the ATP, i. e., 7.25 (Table 2).

DISCUSSION

The lowest number of flies collected in July was due to a cold mass movement in the first two days of sampling, when strong winds (up to 22,4 Km/h) were verified in the area, followed by low temperatures and high relative air humidity.

Our results agree with statements of the local population where the work was carried out, corroborating the fact that the highest number of black flies occur in months of higher precipitation. Cerqueira (1959) in a work developed in the city of Codajás (Middle Solimões River), Amazonas, the largest density of *S. amazonicum* [= *C. argentiscutum*] was also found during the rainy season, considered by him to be from January to June.

Medeiros & Py-Daniel (1999) verified a higher abundance of the species *C. oyapockense*, in Xitei/Xidea, in the Yanomami indigenous area, Roraima, Brazil, in the period of higher precipitation. Shelley (1988) working in Toototobi and Shelley

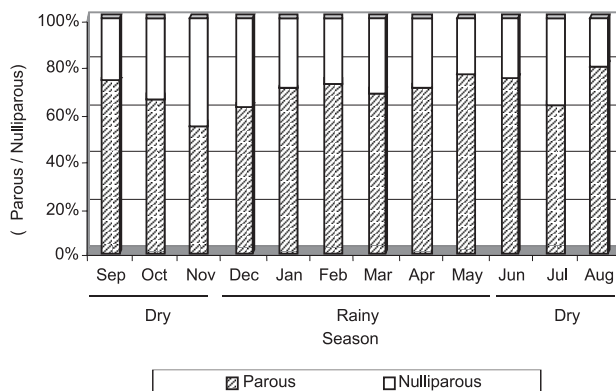


Figure 1 - Distribution of the *Cerqueirellum argentiscutum* females according to the physiologic ovarian condition (parous or nulliparous), collected from September/1999 to August/2000, in Porto Japão, Amazon, Brazil.

et al. (1997), in Catrimani (both Yanomami areas) reported that the highest number of *S. oyapockense* [= *C. oyapockense*] was observed during the rainy period.

The present results provided evidence that *C. argentiscutum* was more abundant in the beginning of the rainy season (from December to February), subsequently decreasing in numbers from March to May, and at the beginning of the dry period (June and July) when the Solimões River is flooded. We observed that from May to July, when the floods reached the forest, the populations of *C. argentiscutum* migrated to the inundated area (also called várzea), where a higher prevalence of the simuliids were found, correlated with a

Table 1 - Seasonal distribution of *Cerqueirellum argentiscutum* biting females in four days of monthly collections, from September/1999 to August/2000, in Porto Japão community, Amazonas, Brazil.

Month/Day	Period	I	II	III	IV	Total	%	Mean/Day±SD n = 04
September/99	Dry	233	284	115	386	1018	6.24	254.5 ±112.6
October	Dry	96	67	136	118	417	2.56	104.3 ±29.7
November	Dry	203	105	116	67	491	3.01	122.8 ±57.4
December	Rainy	342	848	403	172	1765	10.81	441.3 ±288.2
January/00	Rainy	1119	1312	867	789	4087	25.04	1021.8 ±239.3
February	Rainy	847	724	1028	1302	3901	23.90	975.3 ±251.0
March	Rainy	442	364	204	441	1451	8.89	362.8 ±111.9
April	Rainy	284	124	422	326	1156	7.08	289.0 ±124.2
May	Rainy	286	244	428	170	1128	6.91	282.0 ±108.5
June	Dry	103	95	145	142	485	2.97	121.3 ±25.9
July	Dry	12	03	36	41	92	0.56	23.0 ±18.3
August	Dry	109	59	75	86	329	2.02	82.3 ±20.9
Total		4076	4229	3975	4040	16320	100.00	4080.0 ±107.7

Table 2 - Calculated indices of *Cerqueirellum argentiscutum* females collected from September/1999 to August/2000, in Porto Japão community, Amazonas, Brazil.

Parameters	Derivation	Set	Oct	Nov	Dec	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Total
A	Total of Females collected	1018	417	491	1765	4087	3901	1451	1156	1128	485	92	329	16320
	Multiparous Females	270	141	224	664	1227	1064	458	342	266	123	34	66	4879
B	Parous Females	748	276	267	1101	2860	2837	993	814	862	362	58	263	11441
	Females with L1	-	-	-	1	2	2	1	-	-	-	1	1	8
	Females with L2	-	-	-	-	-	1	-	-	-	-	-	-	1
	Females with L2+L3	-	-	-	-	-	1	-	-	-	-	-	-	1
C	Females with L3	-	-	-	-	-	1	-	-	-	-	-	-	1
D	Infected females (mf, L1,L2,L3)	-	-	-	1	2	3	1	-	-	-	1	1	9
E	Infected females (L1,L2,L3)	0	0	0	1	2	3	1	0	0	0	1	1	9
F	No. of L3	0	0	0	0	0	1	0	0	0	0	0	0	1
G	Total number of <i>M. ozzardi</i>	-	-	-	1	3	27	1	-	-	-	1	1	34
H	Number of days in the referred month	30	31	30	31	31	29	31	30	31	30	31	31	366
I	Average number of <i>M. ozzardi</i> infected female	g / d	-	-	1.00	1.50	9.00	1.00	-	-	-	1.00	1	3.78
L	Average number of females man/day	a / 4	254.50	104.25	441.25	1021.75	975.25	362.75	289.00	282.00	121.25	23.00	82.25	340.00
M	Infected females / man / day	d / 4	0.00	0.00	0.25	0.50	0.75	0.25	0.00	0.00	0.00	0.25	0	0.19
N	Infective females / man / day	c / 4	-	-	-	-	0.25	-	-	-	-	-	-	0.25
O	L3 larvae / man / day	Jn	-	-	-	-	1.00	-	-	-	-	-	-	1.00
	PIR1 (Parasitic Infection Rate)	(d/a) x100	0.00	0.00	0.06	0.05	0.08	0.07	0.00	0.00	0.00	1.09	0	0.06
	PIR2	(e/b) x100	0.00	0.00	0.09	0.07	0.11	0.10	0.00	0.00	0.00	1.72	0	0.08
	MBR1 (Monthly Biting Rate)	ah / 4	7635.00	3231.75	13678.75	31674.25	28282.25	11245.25	8670.00	8742.00	3637.50	713.00	2549.75	123742.00
	MBR2	bh / 4	5610.00	2139.00	8532.75	22165.00	20568.25	7695.75	6105.00	6680.50	2715.00	449.50	2038.25	86701.50
	MTP (Monthly Transmission Potential)	(mbr1) f / a	0.00	0.00	0.00	0.00	7.25	0.00	0.00	0.00	0.00	0.00	0.00	7.25
	ATP1													

ATP = Annual Transmission Potential

decrease in the number of specimens sampled alongside the river. It may be suggested that females of this species migrate to the várzea area as a survival strategy, to have access to other sources of blood.

The high number of parous flies collected in this work is probably related to the anthropophilic behavior of *C. argentiscutum* and their high chances of finding a host (man), taking into account the working habits of the local people (setting up agricultural crops at the margins of the river, being exposed daily to the biting activity of this species). Medeiros & Py-Daniel (1999) found 84% of parous female of *C. oyapockense* in Xitei/Xidea; and Shelley *et al.* (1997), collected a higher number of *S. oyapockense* [= *C. oyapockense*] parous females (79.3%) as compared to the nulliparous females (20.7%) in Toototobi.

In a previous report involving several streams in the city of Codajás, Cerqueira (1959) indicated for *S. amazonicum* [= *C. argentiscutum*] a distinctive parasitic infection rate for the various places, totaling 18.5% of the 1367 individuals captured. Shelley & Shelley (1976) found a natural infection rate for *S. amazonicum* [= *C. amazonicum*] estimated in 0.99%, representing 35 individuals out of 3530 collected in humans and bovine cattle, while working in two villages located on the Purus River, close to the municipality of Lábrea, Amazonas. Shelley *et al.* (1980) found *S. amazonicum* [= *C. amazonicum*] and *Simulium* n.sp [= *C. argentiscutum*] naturally infected with *M. ozzardi*, with a parasitic infection rate of 3.1% and 9.7%, respectively (Ticuna tribe, Solimões River, Amazonas).

By comparing the Parasitic Infection Rates (PIR) of the published works cited above, we can say that the PIR values observed in this work are low, particularly in relation to the findings of Cerqueira (1959) and Shelley *et al.* (1980). The low rates (PIR₁ and PIR₂) found in this work are probably due to the absence of mansonellosis in the community of Porto Japão (0% prevalence, in 80% of the examined population). The black flies found infected are most likely to be related to factors such as periodic visits (migrations) of people infected with mansonellosis from other communities located in the Middle and Upper Solimões River, considering the fact that daily movement of ships takes place on the river.

In Brazil, reports on the *M. ozzardi* vectors have not been considering systematic collection (Cerqueira, 1959, Shelley & Shelley, 1976, Shelley *et al.*, 1980, Moraes *et al.*, 1985), and therefore, for comparisons of the Monthly Biting Rate (MBR), we considered some works on species involved in the transmission of the filarial worm *Onchocerca volvulus* (Leuckart) in the indigenous Yanomami area, Roraima, Brazil (Andreazze & Py-Daniel, 1999, Py-Daniel *et al.*, 2000).

Andreazze & Py-Daniel (1999) in the Xitei/Xidea area, determined a MBR₁ of 63.079,7 and a MBR₂ of 37.394,5 for the species *P. incrustata*. Py-Daniel *et al.* (2000) found a MBR₁ of 28.443,0 and a MBR₂ of 18.360,2 in a work conducted in the same place with the species *T. guianense*. Py-Daniel *et al.* (2000) also estimated the MBR for the four

species (*P. incrustata*, *T. guianense*, *C. oyapockense* and *N. exigua*) occurring in Xitei/Xidea, and found a MBR₁ of 92.931,7 and a MBR₂ of 56.936,5.

By comparing the MBRs between these two areas (Porto Japão and Xitei/Xidea), we observed that in Porto Japão, MBRs in only one species was higher than the total MBRs calculated for the four species of Xitei/Xidea area. Therefore, the species *C. argentiscutum*, in some periods of the year, such as January and February, represents a plague for the population living alongside the Solimões River due to the high density of flies and consequently, the discomfort of their bites.

The fact that only one metacyclic larva (L3 stage) was found in February indicates a low ATP for this area, suggesting that a low transmission risk of *M. ozzardi* exists for the population of the study area. According to an index established by the World Health Organization for the vectors of *O. volvulus*, the estimated level in order to maintain transmission is around 100 L3 larvae per person during one year (Davis *et al.*, 1994), which is not yet known for *M. ozzardi* vectors.

ACKNOWLEDGEMENTS

We thank INPA for support, CNPq for the PhD scholarship for the first author, and the Porto Japão community, especially Mr. Aníbal César B. Lima for excellent assistance in the black fly samplings and dissections.

LITERATURE CITED

- Batista, D.; Oliveira, W.R.; Rabello, V.D. 1960. Estudo da patogenicidade da *Mansonella ozzardi* e da sintomatologia da mansonelose. *Revista do Instituto de Medicina Tropical de São Paulo*, 2(5): 281-289.
- Biagi, F.; Tay, J.; Biage, A.M. 1958. Observaciones sobre mansonelosis en la Península de Yucatán. V. *Culicoides furens* como transmissor. *Medicina* 38: 377-379.
- Branco, B.C.; Chamon, W.; Belfort, R.N.; Belfort, Jr.R.; Costa, A.J.A. 1998. Achados oculares entre habitantes do município de Pauini e possível associação entre lesões corneanas e mansonelose na Amazônia. *Arquivo Brasileiro de Oftalmologia*, 61(6): 675-681.
- Buckley, J.J.C. 1934. On the development, in *Culicoides furens* Poey, of *Filaria* (= *Mansonella*) *ozzardi* Manson, 1897. *Journal Helminthologia*, 12: 99-118.
- Cerqueira, N.L. 1959. Sobre a transmissão da *Mansonella ozzardi* 1ª e 2ª notas (Trabalho do Inpa). *Jornal Brasileiro de Medicina*, 1(7): 885-914.
- Davies, J.B.; Crosskey, R.W. 1991. *Simulium* – Vectors of *Onchocerciasis*. World Health Organization, 109 pp.
- Davis, J.R.; Wasserman, S.S.; Trpis, M. 1994. Diurnal biting activity and transmission of *Onchocerca volvulus* (Filariata, Onchocercidae) by *Simulium yabense* (Diptera: Simuliidae) in Liberia. *Journal Medical Entomology*, 31(2): 217-224.

- Deane, M.P. 1949. Sobre a incidência de filárias humanas em Manaus Estado do Amazonas. *Revista do Sesp*, 2(3): 849-858.
- Duke, B.O.L. 1968. Studies on factors influencing the transmission of Onchocerciasis IV. The biting-cycles, infective biting density and transmission potential of 'forest' *Simulium damnosum*. *Annals of Tropical Medicine Parasitology*, 62: 95-106.
- Garrido, C.; Campos, M. 2000. First of presumed parasitic keratitis in indians from the Brazilian Amazon. *Cornea*, 19(6): 817-819.
- Lacerda, N.B.; Rachou, R.G. 1956. Filarioses humanas nas sedes municipais do Estado do Amazonas e dos Territórios do Acre, Guaporé e Rio Branco. *Revista de Malariologia e Doenças Tropicais*, 8(3): 437-442.
- Lowrie, R.C.Jr.; Raccurt, C. 1981. *Mansonella ozzardi* in Haiti II. Arthropod vector studies. *American Journal Tropical Medicine and Hygiene*, 70: 16-17.
- Medeiros, J.F.; Py-Daniel, V. 1999. Atividade hematofágica e infecção natural de três espécies de Simuliidae (Diptera: Culicomorpha) em Xitei/Xidea, área indígena Yanomami, Roraima, Brasil. *Entomologia y Vectores*, 6(3): 210-226.
- Medeiros, J.F.; Py-Daniel, V. 2003. Estado fisiológico e preferência de picadas de *Cerqueirellum argentiscutum* (Shelley & Luna Dias, 1980) (Diptera: Simuliidae), vetor de *Mansonella ozzardi* (Manson, 1897) (Nematoda: Onchocercidae) no Brasil. *Entomologia y Vectores*, 9(4): 505-517.
- Moraes, M.A.P.; Shelley, A.J.; Luna Dias, A.P.A. 1985. *Mansonella ozzardi* no Território Federal de Roraima. Distribuição e achado de um novo vetor na área do rio Surumu. *Memórias do Instituto Oswaldo Cruz*, 80(4): 395-400.
- Nathan, N.B. 1978. *Culicoides pblebotomus*, a vector on *M. ozzardi* in coastal North Trinidad West Indies. *Transaction Royal Society of Tropical Medicine and Hygiene*, 72: 436-437.
- Nathan, M.B.; Tikasingh, E.S.; Munroe, P. 1982. Filariasis in amerindians of western Guyana with observations on transmission of *Mansonella ozzardi* by a *Simulium* species of the *amazonicum* group. *Tropenmedizin Parasitologie*, 33: 219-222.
- Oliveira, W.R. 1961. Filarioses humanas na cidade de Manaus. *O Hospital*, 56(2): 301-303.
- Peterson, J.L.; Bawden, M.P.; Wignell, F.S.; Latorre, C.R.; Johnson, C.M.; Miranda, C. 1984. *Mansonella ozzardi* en el Darien (Panama). *Revista Médica Panama*, 9: 236-246.
- Py-Daniel, V.; Andrezza, R.; Medeiros, J.F. 2000. Projeto Piloto Xitei/Xidea, (Roraima). I - índices epidemiológicos da transmissão de *Onchocerca volvulus* (Leuckart, 1893) para os anos de 1995-1996. *Entomologia y Vectores*, 7(4): 389-444.
- Py-Daniel, V.; Moreira-Sampaio RT 1994. *Jalacingomyia* Gen. (Culicomorpha); A ressurreição de Gymnopauidinae; a eliminação do nível tribal, Apresentação de novos caracteres e a redescoberta dos estágios larval e pupal de *Simulium colombaschense* (Fabricius, 1787) (Diptera, Simuliidae). *Memorias del Caicet*, IV(1,2): 101-148.
- Ramírez-Perez, J. 1977. *Estudo sobre la morfologia de Simulium metallicum vector de la Onchocercosis humana en Venezuela*. Organizacion Panamericana de la Salud/Organizacion Mundial de la salud, Publicación Científica N° 338, iii + 1 -140 pp.
- Ribeiro, M.N.G.; Adis, J. 1984. Local rainfall variability - a potential bias for bioecological studies in the Central Amazon. *Acta Amazonica*, 14(1-2): 159-174.
- Shelley, A.J. 1988. Vector aspects of the epidemiology of Onchocerciasis in Latin America. *Annual Review of Entomology* 30: 337-336.
- Shelley, A.J.; Coscarón, S. 2001. Simuliid blackflies (Diptera: Simuliidae) and ceratopogonid Midges (Diptera: Ceratogonidae) as vectors of *Mansonella ozzardi* (Nematoda: Onchocercidae) in northern Argentina. *Memórias do Instituto Oswaldo Cruz* 96(4): 451-458.
- Shelley, A.J.; Lowry, C.A.; Maia-Herzog, M.; Luna Dias, A.P.A.; Moraes, M.A.P. 1997. Biosystematic studies on the Simuliidae (Diptera) of the Amazonia Onchocerciasis focus. *Bull British Museum (Natural History) Entomology*, 66(1): 1-121.
- Shelley, A.J.; Luna Dias, A.P.A.; Moraes, M.A.P. 1980. *Simulium* species of the *amazonicum* group as vectors de *Mansonella ozzardi* in the Brazilian Amazon. *Trans Royal Society of Tropical Medicine Hygiene*, 74: 784-788.
- Shelley, A.J.; Shelley, A. 1976. Further evidence for the transmission of *Mansonella ozzardi* by *Simulium amazonicum* in Brazil. *Annals Tropical Medical Parasitology*, 70: 213-217.
- Tavares, A.M.; Fraiha Neto, H. 1997. Mansonelose. In: Queiroz de Leão, R.N.(Ed). *Doenças Infecciosas e Parasitárias, Enfoque Amazônico*, p. 733-737.
- Tidwell, M.A.; Tidwell, M.A.; Muñoz de Hoyos, P. 1980. Development of *Mansonella ozzardi* in blackflies of the *Simulium sanguineum* group from eastern Vaupés, Colombia. *American Journal of Tropical Medicine and Hygiene*, 29: 1209-1214.
- Tidwell, M.A.; Tidwell, M.A. 1982. Development of *Mansonella ozzardi* in *Simulium amazonicum*, *S. argentiscutum* and *Culicoides insinuatus* from Amazonas, Colombia. *American Journal Tropical Medicine Hygiene*, 3: 1137-1141.
- Yazarbal, L.; Basáñez, M.G.; Ramírez-Pérez, J.; Ramírez, A.; Botto, C.; Yazarbal, A. 1985. Experimental and natural infection of *Simulium sanchezi* by *Mansonella ozzardi* in the middle Orinoco region of Venezuela. *Transactions Royal Society of Tropical Medicine Hygiene*, 79: 29-33.

RECEBIDO EM 23/01/2004

ACEITO EM 19/04/2004

